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(54) Title: METHODS OF TREATING ORTHOMYXOVIRAL INFECTIONS

(55) Abstract: Provided are novel iminosugars and methods of treating and/or preventing a disease or condition caused by or associated with a virus belonging to the Orthomyxoviridae family using iminosugars, such as DNJ derivatives.

FIGURE 7: UV-4 THERAPEUTIC MODE VS. INFLUENZA H1N1
METHODS OF TREATING ORTHOMYXOVIRAL INFECTIONS

RELATED APPLICATIONS

The present application claims priority to a) US provisional application no. 61/272,254 filed September 4, 2009; b) US provisional application no. 61/282,508 filed February 22, 2010 and c) US provisional application no. 61/353,935 filed June 11, 2010, each of which is incorporated herein by reference in its entirety.

FIELD

The present application relates to iminosugars and methods of treating viral infections with iminosugars and, in particular, to the use of iminosugars for treatment and/or prevention of viral infections caused by or associated with a virus belonging to the Orthomyxoviridae family.

SUMMARY

One embodiment is a method of treating or preventing a disease or condition caused by or associated with a virus belonging to the Orthomyxoviridae family, which method comprises administering to a subject in need thereof an effective amount of a compound of the formula:

\[
\begin{align*}
W_1 & \quad \text{W}_{2,\text{o}} \\
\quad & \quad \text{W}_{3,\text{O}} \\
\quad & \quad \text{W}_{4,\text{O}} \\
\end{align*}
\]

, or a pharmaceutically acceptable salt thereof, wherein R is either selected from substituted or unsubstituted alkyl groups, substituted or unsubstituted cycloalkyl groups, substituted or unsubstituted aryl groups, or substituted or unsubstituted oxaalkyl groups; or wherein R is
R₁ is a substituted or unsubstituted alkyl group;
X₁₋₅ are independently selected from H, NO₂, N₃, or NH₂;
Y is absent or is a substituted or unsubstituted C₁-alkyl group, other than carbonyl; and
Z is selected from a bond or NH; provided that when Z is a bond, Y is absent, and provided that when Z is NH, Y is a substituted or unsubstituted C₁-alkyl group, other than carbonyl; and
wherein W₁₋₄ are independently selected from hydrogen, substituted or unsubstituted alkyl groups, substituted or unsubstituted haloalkyl groups, substituted or unsubstituted alkanoyl groups, substituted or unsubstituted aroyl groups, or substituted or unsubstituted haloalkanoyl groups.
Another embodiment is a method of inhibiting infectivity of a cell infected with a virus belonging to the Orthomyxoviridae family, which method comprises contacting a cell infected with a virus belonging to the Orthomyxoviridae family with an effective amount of a compound of the formula:

, or a pharmaceutically acceptable salt thereof, wherein R is either selected from substituted or unsubstituted alkyl groups, substituted or unsubstituted cycloalkyl groups, substituted or unsubstituted aryl groups, or substituted or unsubstituted oxaalkyl groups; or wherein R is
$R_1$ is a substituted or unsubstituted alkyl group;

$X_{1-5}$ are independently selected from H, NO$_2$, N$_3$, or NH$_2$;

Y is absent or is a substituted or unsubstituted C$_1$-alkyl group, other than carbonyl; and

Z is selected from a bond or NH; provided that when Z is a bond, Y is absent, and provided

that when Z is NH, Y is a substituted or unsubstituted C$_1$-alkyl group, other than carbonyl;

and

wherein $W_{1-4}$ are independently selected from hydrogen, substituted or unsubstituted alkyl
groups, substituted or unsubstituted haloalkyl groups, substituted or unsubstituted alkanoyl
groups, substituted or unsubstituted aroyl groups, or substituted or unsubstituted haloalkanoyl
groups.

**DRAWINGS**

Figures 1(A)-(E) present chemical formulas of the following iminosugars: A) $N$-Butyl
deoxynojirimycin (NB-DNJ or UV-1); B) $N$-Nonyl deoxynojirimycin (NN-DNJ or UV-2); C)
$N$-(7-Oxadecyl)deoxynojirimycin (N7-O-DNJ or UV-3); D) $N$-(9-Methoxynonyl)
deoxynojirimycin (N9-DNJ or UV-4); E) $N$-(9-{4'-azido-2'-nitrophenyl}-6-
aminoheptyl)deoxynojirimycin (NAP-DNJ or UV-5).

Figure 2 is a synthesis scheme for NN-DNJ.

Figures 3A-D illustrate synthesis of N7-O-DNJ. In particular, Figure 3A shows a sequence
of reactions leading to N7-O-DNJ; Figure 3B illustrates preparation of 6-propyloxy-1-
hexanol; Figure 3C illustrates preparation of 6-propyloxy-1-hexanal; Figure 3D illustrates
synthesis of N7-O-DNJ.

Figures 4A-C relate to synthesis of $N$-(9-Methoxynonyl) deoxynojirimycin. In particular,
Figure 4A illustrates preparation of 9-methoxy-1-nonanol; Figure 4B illustrates preparation
of 9-methoxy-1-nonanal; Figure 4C illustrates synthesis of $N$-(9-Methoxynonyl)
deoxynojirimycin.
Figure 5 presents effects of 10 day administration of UV-5 on survival of mice infected with influenza A H1N1.

Figure 6 presents *in vivo* safety data for UV-4 and UV-5.

Figure 7 presents survival data after H1/N1 virus challenge for mice treated with UV-4 versus control mice.

**DETAILED DESCRIPTION**

Related Patent Documents

The following patent documents, which are all incorporated herein by reference in their entirety, may be useful for understanding the present disclosure:

1) US patent no. 6,545,021;
2) US patent no. 6,809,803;
3) US patent no. 6,689,759;
4) US patent no. 6,465,487;
5) US patent no. 5,622,972;
6) US patent application no. 12/656,992 filed February 22, 2010;
7) US patent application no. 12/656,993 filed February 22, 2010;
8) US patent application no. 12/813,882 filed June 11, 2010;
9) US patent provisional application no. 61/282,507 filed February 22, 2010;
10) US patent provisional application no. 61/272,252 filed September 4, 2009;
11) US provisional application no. 61/272,253 filed September 4, 2009;
12) US provisional application no. 61/272,254 filed September 4, 2009;
13) US provisional application no. 61/282,508 filed February 22, 2010;

**Definition of terms**

Unless otherwise specified, “a” or “an” means “one or more.”

As used herein, the term “viral infection” describes a diseased state, in which a virus invades a healthy cell, uses the cell’s reproductive machinery to multiply or replicate and ultimately lyse the cell resulting in cell death, release of viral particles and the infection of other cells by
the newly produced progeny viruses. Latent infection by certain viruses is also a possible result of viral infection.

As used herein, the term “treating or preventing viral infection” means to inhibit the replication of the particular virus, to inhibit viral transmission, or to prevent the virus from establishing itself in its host, and to ameliorate or alleviate the symptoms of the disease caused by the viral infection. The treatment is considered therapeutic if there is a reduction in viral load, decrease in mortality and/or morbidity.

IC50 or IC90 (inhibitory concentration 50 or 90) is a concentration of a therapeutic agent, such as an iminosugar, used to achieve 50% or 90% reduction of viral load, respectively.

Disclosure

The present inventors discovered that certain iminosugars, such as deoxynojirimycin derivatives, can be effective against viruses belonging to the Orthomyxoviridae family, also known as orthomyxoviruses.

In particular, iminosugars can be useful for treating and/or preventing a disease or condition caused by or associated with a virus belonging to the Orthomyxoviridae family.

The Orthomyxoviridae family is a family of RNA viruses that includes five genera: Influenzavirus A, Influenzavirus B, Influenzavirus C, Isavirus and Thogotovirus. The first three genera contain viruses that can cause influenza in vertebrates, including birds, humans and other mammals.

The Influenzavirus A genus includes a single species, which can causes influenza in birds and certain mammals, including humans, pigs, felines, canines and equines.

Influenza A viruses are negative sense, single-stranded, segmented RNA viruses. Several subtypes of Influenza A virus exist, labeled according to an H number (for the type of hemagglutinin) and an N number (for the type of neuraminidase). Currently known 16 different H antigens (H1 to H16) and nine different N antigens (N1 to N9). Serotypes and subtypes of Influenza A include H1N1 Influenza A; H1N2 Influenza A; H2N2 Influenza A; H3N1 Influenza A; H3N2 Influenza A; H3N8 Influenza A; H5N1 Influenza A; H5N2 Influenza A; H5N3 Influenza A; H5N8 Influenza A; H5N9 Influenza A; H5N9 Influenza A; H7N1 Influenza A; H7N2 Influenza A; H7N3 Influenza A; H7N4 Influenza A; H7N7 Influenza A; H7N2 Influenza A; H10N7 Influenza A.
The Influenzavirus B genus includes a single species, which can cause influenza in humans and seals.

The Influenzavirus C genus includes a single species, which can cause influenza in humans and pigs.

In many embodiments, the iminosugar may be N-substituted deoxyojirimycin. In some embodiments, such N-substituted deoxyojirimycin may be a compound of the following formula:

![Chemical structure](image)

where \(W_{1,4}\) are independently selected from hydrogen, substituted or unsubstituted alkyl groups, substituted or unsubstituted haloalkyl groups, substituted or unsubstituted alkanoyl groups, substituted or unsubstituted aryl groups, or substituted or unsubstituted haloalkanoyl groups.

In some embodiments, R may be selected from substituted or unsubstituted alkyl groups, substituted or unsubstituted cycloalkyl groups, substituted or unsubstituted aryl groups, or substituted or unsubstituted oxaalkyl groups.

In some embodiments, R may be substituted or unsubstituted alkyl groups and/or substituted or unsubstituted oxaalkyl groups comprise from 1 to 16 carbon atoms, from 4 to 12 carbon atoms or from 8 to 10 carbon atoms. The term “oxaalkyl” refers to an alkyl derivative, which can contain from 1 to 5 or from 1 to 3 or from 1 to 2 oxygen atoms. The term “oxaalkyl” includes hydroxyterminated and methoxyterminated alkyl derivatives.

In some embodiments, R may be selected from, but is not limited to \(-(CH_2)_kOCH_3,\)
\n\[-(CH_2)_kOCH_2CH_3, -(CH_2)_kO(CH_2)_2CH_3, -(CH_2)_kO(CH_2)_3CH_3, -(CH_2)_2O(CH_2)_3CH_3,\]
\n\[-(CH_2)_2O(CH_2)_3CH_3; -(CH_2)_2O(CH_2)_2CH_3; -(CH_2)_2OH; -(CH_2)_2OCH_3.\]

In some embodiments, R may be branched or unbranched, substituted or unsubstituted alkyl group. In certain embodiments, the alkyl group may be a long chain alkyl group, which may
be C6-C20 alkyl group; C8-C16 alkyl group; or C8-C10 alkyl group. In some embodiments, R may be a long chain oxaalkyl group, i.e. a long chain alkyl group, which can contain from 1 to 5 or from 1 to 3 or from 1 to 2 oxygen atoms.

In some embodiments, R may have the following formula

\[ \text{X}_1 - \text{Y} - \text{Z} - \text{X}_3 \]

, where \( \text{R}_1 \) is a substituted or unsubstituted alkyl group;

\( \text{X}_{1-5} \) are independently selected from H, NO\(_2\), N\(_3\), or NH\(_2\);

Y is absent or is a substituted or unsubstituted C\(_1\)-alkyl group, other than carbonyl; and

Z is selected from a bond or NH; provided that when Z is a bond, Y is absent, and provided that when Z is NH, Y is a substituted or unsubstituted C\(_1\)-alkyl group, other than carbonyl.

In some embodiments, Z is NH and \( \text{R}_1\)-Y is a substituted or unsubstituted alkyl group, such as C2-C20 alkyl group or C4-C12 alkyl group or C4-C10 alkyl group.

In some embodiments, \( \text{X}_1 \) is NO\(_2\) and \( \text{X}_3 \) is N\(_3\). In some embodiments, each of \( \text{X}_2, \text{X}_4 \) and \( \text{X}_5 \) is hydrogen.

In some embodiments, the iminosugar may be a DNJ derivative disclosed in U.S. Patent application publication no. 2007/0275998, which is incorporated herein by reference.

In some embodiments, the iminosugar may be one of the compounds presented in Figure 1. Methods of synthesizing deoxyojirimycin derivatives are disclosed, for example, in U.S. Patent Nos. 5,622,972, 5,200,523, 5,043,273, 4,994,572, 4,246,345, 4,266,025, 4,405,714, and 4,806,650 and U.S. Patent application publication no. 2007/0275998, which are all incorporated herein by reference.

In some embodiments, the iminosugar may be in a form of a salt derived from an inorganic or organic acid. Pharmaceutically acceptable salts and methods for preparing salt forms are disclosed, for example, in Berge et al. (J. Pharm. Sci. 66:1-18, 1977). Examples of appropriate salts include but are not limited to the following salts: acetate, adipate, alginate, citrate, aspartate, benzoate, benzenesulfonate, bisulfate, butyrate, camphorate, camphorsulfonate, digluconate, cyclopentanepropionate, dodecylsulfate, ethanesulfonate,
glucoheptanoate, glycerophosphate, hemisulfate, heptanoate, hexanoate, fumarate, hydrochloride, hydrobromide, hydroiodide, 2-hydroxyethanesulfonate, lactate, maleate, methanesulfonate, nicotinate, 2-naphthalenesulfonate, oxalate, palmoate, pectinate, persulfate, 3-phenylpropionate, picrate, pivalate, propionate, succinate, tartrate, thiocyanate, tosylate, mesylate, and undecanoate.

In some embodiments, the iminosugar may also be used in a form of a prodrug. Prodrugs of DNJ derivatives, such as the 6-phosphorylated DNJ derivatives, are disclosed in U.S. Patents nos. 5,043,273 and 5,103,008.

In some embodiments, the iminosugar may be used as a part of a composition, which further comprises a pharmaceutically acceptable carrier and/or a component useful for delivering the composition to an animal. Numerous pharmaceutically acceptable carriers useful for delivering the compositions to a human and components useful for delivering the composition to other animals such as cattle are known in the art. Addition of such carriers and components to the composition of the invention is well within the level of ordinary skill in the art.

In some embodiments, the pharmaceutical composition may consist essentially of N-substituted deoxyxojirimycin, which may mean that the N-substituted deoxyxojirimycin is the only active ingredient in the composition.

Yet in some embodiments, N-substituted deoxyxojirimycin may be administered with one or more additional antiviral compounds.

In some embodiments, the treatment or prevention of the disease or condition caused by or associated with a virus belonging to the Orthomyxoviridae family may be performed without administering N-(phosphonoacetyl)-L-aspartic acid to the subject, to whom the iminosugar is being administered. N-(phosphonoacetyl)-L-aspartic acid is disclosed, for example, in U.S. patent no. 5,491,135.

In some embodiments, the treatment or prevention of the disease or condition caused by or associated with a virus belonging to the Orthomyxoviridae family may be performed without administering to the subject a pyrroliidine compound, such as compounds disclosed in U.S. patent no. 5,021,427 and U.S. patent publication 20070155814.
In some embodiments, the treatment or prevention of the disease or condition caused by or associated with a virus belonging to the Orthomyxoviridae family may be performed without administering to the subject australine.

In some embodiments, the iminosugar, such as N-substituted deoxynojirimycin, may be used in a liposome composition, such as those disclosed in US publications nos. 2008/0138351 and 2009/0252785 as well as in US application No. 12/732630 filed March 26, 2010. The iminosugar, such as N-substituted DNJ derivative, may be administered to a cell or an animal affected by a virus. The iminosugar may inhibit morphogenesis of the virus, or it can treat the individual. The treatment may reduce, abate, or diminish the virus infection in the animal.

Animals that can be infected with a virus that belongs to the Orthomyxoviridae family, include vertebrates, such as birds and mammals, including primates, such as humans; felines; equines, and canines.

The amount of iminosugar administered to an animal or to an animal cell to the methods of the invention may be an amount effective to inhibit the morphogenesis of a virus belonging to the Orthomyxoviridae family from the cell. The term "inhibit" as used herein may refer to the detectable reduction and/or elimination of a biological activity exhibited in the absence of the iminosugar. The term "effective amount" may refer to that amount of the iminosugar necessary to achieve the indicated effect. The term "treatment" as used herein may refer to reducing or alleviating symptoms in a subject, preventing symptoms from worsening or progressing, inhibition or elimination of the causative agent, or prevention of the infection or disorder related to the virus belonging to the Orthomyxoviridae family in a subject who is free therefrom.

Thus, for example, treatment of the disease caused by or associated with a virus may include destruction of the infecting agent, inhibition of or interference with its growth or maturation, and neutralization of its pathological effects. The amount of the iminosugar which may be administered to the cell or animal is preferably an amount that does not induce toxic effects which may outweigh the advantages which accompany its administration.

Actual dosage levels of active ingredients in the pharmaceutical compositions may vary so as to administer an amount of the active compound(s) that is effective to achieve the desired therapeutic response for a particular patient.
The selected dose level can depend on the activity of the iminosugar, the route of administration, the severity of the condition being treated, and the condition and prior medical history of the patient being treated. However, it is within the skill of the art to start doses of the compound(s) at levels lower than required to achieve the desired therapeutic effect and to gradually increase the dosage until the desired effect is achieved. If desired, the effective daily dose may be divided into multiple doses for purposes of administration, for example, two to four doses per day. It will be understood, however, that the specific dose level for any particular patient can depend on a variety of factors, including the body weight, general health, diet, time and route of administration and combination with other therapeutic agents and the severity of the condition or disease being treated. The adult human daily dosage may range from between about one microgram to about one gram, or from between about 10 mg and 100 mg, of the iminosugar per 10 kilogram body weight. In some embodiments, a total daily dose may be from 0.1 mg/kg body weight to 100 mg/kg body weight or from 1 mg/kg body weight to 60 mg/kg body weight or from 2 mg/kg body weight to 50 mg/kg body weight or from 3 mg/kg body weight to 30 mg/kg body weight. The daily dose may be administered over one or more administering events over day. For example, in some embodiments, the daily dose may be distributed over two (BID) administering events per day, three administering events per day (TID) or four administering events (QID). In certain embodiments, a single administering event dose ranging from 1 mg/kg body weight to 10 mg/kg body weight may be administered BID or TID to a human making a total daily dose from 2 mg/kg body weight to 20 mg/kg body weight or from 3 mg/kg body weight to 30 mg/kg body weight. Of course, the amount of the iminosugar which should be administered to a cell or animal can depend upon numerous factors well understood by one of skill in the art, such as the molecular weight of the iminosugar and the route of administration. Pharmaceutical compositions that are useful in the methods of the invention may be administered systemically in oral solid formulations, ophthalmic, suppository, aerosol, topical or other similar formulations. For example, it may be in the physical form of a powder, tablet, capsule, lozenge, gel, solution, suspension, syrup, or the like. In addition to the iminosugar, such pharmaceutical compositions may contain pharmaceutically-acceptable carriers and other ingredients known to enhance and facilitate drug administration. Other possible formulations, such as nanoparticles, liposomes, resealed erythrocytes, and
immunologically based systems may also be used to administer the iminosugar. Such pharmaceutical compositions may be administered by a number of routes. The term "parenteral" used herein includes subcutaneous, intravenous, intraarterial, intrathecal, and injection and infusion techniques, without limitation. By way of example, the pharmaceutical compositions may be administered orally, topically, parenterally, systemically, or by a pulmonary route.

These compositions may be administered a in a single dose or in multiple doses which are administered at different times. Because the inhibitory effect of the composition upon a virus belonging to the Orthomyxoviridae family may persist, the dosing regimen may be adjusted such that virus propagation is retarded while the host cell is minimally effected. By way of example, an animal may be administered a dose of the composition of the invention once per week, whereby virus propagation is retarded for the entire week, while host cell functions are inhibited only for a short period once per week.

Embodiments described herein are further illustrated by, though in no way limited to, the following working examples.

Working Examples

1. Synthesis of N-Nonyl DNJ

Table 1. Materials for NN-DNJ synthesis

<table>
<thead>
<tr>
<th>Name</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>DNJ</td>
<td>500 mg</td>
</tr>
<tr>
<td>Nonanal</td>
<td>530 mg</td>
</tr>
<tr>
<td>Ethanol</td>
<td>100 mL</td>
</tr>
<tr>
<td>AcOH</td>
<td>0.5 mL</td>
</tr>
<tr>
<td>Pd/C</td>
<td>500 mg</td>
</tr>
</tbody>
</table>

Procedure: A 50-mL, one-necked, round-bottom flask equipped with a magnetic stirrer was charged with DNJ (500 mg), ethanol (100 mL), nonanal (530 mg), and acetic acid (0.5 mL) at room temperature. The reaction mixture was heated to 40-45 °C and stirred for 30-40 minutes under nitrogen. The reaction mixture was cooled to ambient temperature and Pd/C
was added. The reaction flask was evacuated and replaced by hydrogen gas in a balloon. This process was repeated three times. Finally, the reaction mixture was stirred at ambient temperature overnight. The progress of reaction was monitored by TLC (Note 1). The reaction mixture was filtered through a pad of Celite and washed with ethanol. The filtrate was concentrated in vacuo to get the crude product. The crude product was purified by column chromatography (230-400 mesh silica gel). A solvent gradient of methanol in dichloromethane (10-25%) was used to elute the product from the column. All fractions containing the desired product were combined, and concentrated in vacuo to give the pure product (420mg). Completion of the reaction was monitored by thin layer chromatography (TLC) using a thin layer silica gel plate; eluent; methanol : dichloromethane = 1:2

2. Synthesis of N-7-Oxadecyl DNJ

2a. Synthesis of 6-propyloxy-1-hexanol

Table 2. Materials for synthesis of 6-propyloxy-1-hexanol

<table>
<thead>
<tr>
<th>Name</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>1,6-hexanediol</td>
<td>6.00 g</td>
</tr>
<tr>
<td>1-Iodopropane</td>
<td>8.63 g</td>
</tr>
<tr>
<td>Potassium tert-butoxide</td>
<td>5.413 mg</td>
</tr>
<tr>
<td>THF</td>
<td>140 mL</td>
</tr>
</tbody>
</table>

Procedure: a 500-mL, one-necked, round-bottom flask equipped with a magnetic stirrer was charged with 1,6-hexanediol (6.00 g), potassium tert-butoxide (5.413 g) at room temperature. The reaction mixture was stirred for one hour, and then 1-iodopropane (8.63 g) was added. The reaction mixture was heated to 70-80 °C and stirred overnight. The progress of reaction was monitored by TLC (Note 1). After completion of the reaction, water was added to the reaction mixture, and extracted with ethyl acetate (2 x 100 mL). The combined organic layers were concentrated in vacuo to get the crude product. The crude product was dissolved in dichloromethane and washed with water, and then brine, dried over sodium sulfate. The organic layer was concentrated in vacuo to get the crude product. The crude product was purified by column chromatography using 230-400 mesh silica gel. A solvent gradient of
ethyl acetate in hexanes (10-45%) was used to elute the product from the column. All fractions containing the desired pure product were combined and concentrated *in vacuo* to give pure 6-propyloxy-1-hexanol (lot D-1029-048, 1.9 g, 25%). Completion of the reaction was monitored by thin layer chromatography (TLC); (eluent: 60% ethyl acetate in hexanes).

2b. Preparation of 6-propyloxy-1-hexanal

Table 3. Materials for preparation of 6-propyloxy-1-hexanal

<table>
<thead>
<tr>
<th>Name</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>6-Propyloxy-1-hexanol</td>
<td>1.00 g</td>
</tr>
<tr>
<td>PDC</td>
<td>4.70 g</td>
</tr>
<tr>
<td>Celite</td>
<td>1.00 g</td>
</tr>
<tr>
<td>NaOAc</td>
<td>100 mg</td>
</tr>
<tr>
<td>CH$_2$Cl$_2$</td>
<td>10 mL</td>
</tr>
</tbody>
</table>

Procedure: a 50-mL, one-necked, round-bottom flask equipped with a magnetic stirrer was charged with 6-propyloxy-1-hexanol (1.0 g), PDC (4.7 g), dichloromethane (10 mL), Celite (1.0 g), and sodium acetate (100 mg). The reaction mixture was stirred at room temperature under nitrogen for 5 minutes. PDC (4.70 g) was added to the reaction mixture, and stirred overnight. The progress of reaction was monitored by TLC (Note 1). After completion of the reaction, the reaction mixture was directly loaded on the column (230-400 mesh silica gel). A solvent gradient of dichloromethane in ethyl acetate (10-20%) was used to elute the product from the column. All fractions containing the desired pure product were combined and concentrated *in vacuo* to give pure 6-propyloxy-1-hexanal (lot D-1029-050, 710 mg, 71%). Completion of the reaction was monitored by thin layer chromatography (TLC); (eluent: 60% ethyl acetate in hexanes).

2c Synthesis of N-7-Oxadecyl-DNJ

Table 4. Materials for Synthesis of N-7-Oxadecyl-DNJ

<table>
<thead>
<tr>
<th>Name</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>DNJ</td>
<td>500 mg</td>
</tr>
</tbody>
</table>
Table 5. Materials for preparation of 9-methoxy-1-nonanol

<table>
<thead>
<tr>
<th>Name</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>1,9-nonanediol</td>
<td>10.0 g</td>
</tr>
<tr>
<td>Dimethyl sulfate</td>
<td>41.39 g</td>
</tr>
<tr>
<td>Sodium hydroxide</td>
<td>5.0 g</td>
</tr>
<tr>
<td>DMSO</td>
<td>100 mL</td>
</tr>
</tbody>
</table>

Procedure: a 50-mL, one-necked, round-bottom flask equipped with a magnetic stirrer was charged with DNJ (500 mg), ethanol (15 mL), 6-propyloxy-1-hexanal (585 mg), and acetic acid (0.1mL) at room temperature. The reaction mixture was heated to 40-45 °C and stirred for 30-40 minutes under nitrogen. The reaction mixture was cooled to ambient temperature and Pd/C was added. The reaction flask was evacuated and replaced by hydrogen gas in a balloon. This process was repeated three times. Finally, the reaction mixture was stirred at ambient temperature overnight. The progress of reaction was monitored by TLC (Note 1). The reaction mixture was filtered through a pad of Celite and washed with ethanol. The filtrate was concentrated in vacuo to get the crude product. The crude product was purified by column chromatography (230-400 mesh silica gel). A solvent gradient of methanol in dichloromethane (10-40%) was used to elute the product from the column. All fractions containing the desired product were combined, and concentrated in vacuo to give the pure product. (Lot: D-1029-052 (840 mg). Completion of the reaction was monitored by thin layer chromatography (TLC); (eluent: 50% methanol in dichloromethane).

3. Synthesis of N-(9-methoxy)-nonyl DNJ

3a Preparation of 9-methoxy-1-nonanol
Procedure: a 500-mL, one-necked, round-bottom flask equipped with a magnetic stirrer and stir bar was charged with 1,9-nonanediol (10.00 g, 62.3 mmol) in dimethyl sulfoxide (100 mL) and \( \text{H}_2\text{O} \) (100 mL). To this was added slowly a solution of sodium hydroxide (5.0 g, 125.0 mmol) in \( \text{H}_2\text{O} \) (10 mL) at room temperature. During addition of sodium hydroxide the reaction mixture generated heat and the temperature rose to \( \sim 40^\circ \text{C} \). The mixture was stirred for one hour, and then dimethyl sulfate (16.52 g, 131 mmol) was added in four portions while maintaining the temperature of the reaction mixture at \( \sim 40^0 \text{C} \). The reaction mixture was stirred at room temperature overnight. Progress of the reaction was monitored by TLC (Note 1). TLC monitoring indicated that the reaction was 25 % conversion. At this stage additional dimethyl sulfate (24.78g, 196.44 mmol) was added and the resulting mixture was stirred at room temperature for an additional 24 h. After completion of the reaction, sodium hydroxide (10% solution in water) was added to the reaction mixture to adjust the pH of the solution to 11-13. The mixture was stirred at room temperature for 2 h and extracted with dichloromethane (3 x 100 mL). The combined organic layers were washed with \( \text{H}_2\text{O} \) (200 mL), brine (150 mL), dried over anhydrous sodium sulfate (20 g), filtered and concentrated in vacuo to obtain a crude product (14 g). The crude product was purified by column chromatography using 250-400 mesh silica gel. A solvent gradient of ethyl acetate in hexanes (10-50%) was used to elute the product from the column. All fractions containing the desired pure product were combined and concentrated in vacuo to give pure 9-methoxy-1-nonanol (lot D-1027-155, 2.38 g, 21.9 %). Completion of the reaction was monitored by thin layer chromatography (TLC) using a thin layer silica gel plate; eluent: 60% ethyl acetate in hexanes.

3b Preparation of 9-methoxy-1-nonanol

Table 6. Materials for preparation of 9-methoxy-1-nonanol

<table>
<thead>
<tr>
<th>Name</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>9-methoxy-1-nonanol</td>
<td>1.0 g</td>
</tr>
<tr>
<td>PDC</td>
<td>4.7 g</td>
</tr>
<tr>
<td>Molecular sieves, 3A</td>
<td>1.0 g</td>
</tr>
<tr>
<td>NaOAc</td>
<td>0.1 g</td>
</tr>
<tr>
<td>( \text{CH}_2\text{Cl}_2 )</td>
<td>10 mL</td>
</tr>
</tbody>
</table>
Procedure: a 50-mL, one-necked, round-bottom flask equipped with a magnetic stirrer and stir bar was charged with 9-methoxy-nonanol (1.0 g, 5.9 mmol), dichloromethane (10 mL), molecular sieves (1.0 g, 3A), sodium acetate (0.1 g) at room temperature. The reaction mixture was stirred at room temperature under nitrogen for 5 minutes. The reaction mixture was charged with pyridinium dichromate (4.7 g, 12.5 mmol) and stirred overnight. The progress of reaction was monitored by TLC (Note 1). After completion of the reaction, the reaction mixture was filtered through a bed of silica gel (~15 g). The filtrate was evaporated in vacuo to obtain a crude compound. This was purified by column chromatography using silica gel column (250-400 mesh, 40 g). A solvent gradient of ethyl acetate in hexane (10-50%) was used to elute the product from the column. All fractions containing the desired pure product were combined and concentrated in vacuo to give pure 9-methoxy-nonanal (lot D-1027-156, 553 mg, 54.4%). Completion of the reaction was monitored by thin layer chromatography (TLC) using a thin layer silica gel plate; eluent: 60% ethyl acetate in hexanes.

3c Synthesis of N-(9-methoxy)-nonyl DNJ

Table 7. Materials for synthesis of N-(9-methoxy)-nonyl DNJ

<table>
<thead>
<tr>
<th>Name</th>
<th>Amount</th>
</tr>
</thead>
<tbody>
<tr>
<td>DNJ</td>
<td>300 mg</td>
</tr>
<tr>
<td>9-methoxy-1-nonanal</td>
<td>476 mg</td>
</tr>
<tr>
<td>Pd/C</td>
<td>200 mg</td>
</tr>
<tr>
<td>Ethanol</td>
<td>20 mL</td>
</tr>
</tbody>
</table>

Procedure: a 50-mL, two-necked, round-bottom flask equipped with magnetic stirrer and a stir bar was charged with DNJ (300 mg, 1.84 mmol), ethanol (20 mL), 9-methoxy-1-nonanal (476 mg, 2.76 mmol) at room temperature. The reaction mixture was stirred for 5-10 minutes under nitrogen and Pd/C was added at room temperature. The reaction mixture was evacuated and was replaced by hydrogen gas using a balloon. This process was repeated three times and then reaction mixture was stirred under atmospheric hydrogen at room temperature. The
progress of reaction was monitored by TLC (Note 1). The reaction mixture was filtered through a bed of Celite and was washed with ethanol (20 mL). The filtrate was concentrated in vacuo to get a crude product. The crude product was purified by column chromatography using 250-400 mesh silica gel (20 g). A solvent gradient of methanol in ethyl acetate (5-25%) was used to elute the product from the column. All fractions containing the desired pure product were combined, and concentrated in vacuo to give an off white solid. The solid was triturated in ethyl acetate (20 mL), filtered and dried in high vacuum to give a white solid [lot: D-1027-158 (165.3 mg, 28.1%). Completion of the reaction was monitored by thin layer chromatography (TLC) using a thin layer silica gel plate; eluent: 50% methanol in dichloromethane.

4. Effects of iminosugars against Influenza A virus

Table provides data for inhibition of infectivity of Influenza A virus H3N2 (Hong Kong) for NB-DNJ (UV-1), NN-DNJ (UV-2), N7-O-DNJ (UV-3), N9-DNJ (UV-4) and NAP-DNJ (UV-5).

<table>
<thead>
<tr>
<th>Compound</th>
<th>IC90, μM</th>
</tr>
</thead>
<tbody>
<tr>
<td>UV-1</td>
<td>20</td>
</tr>
<tr>
<td>UV-2</td>
<td>0.2</td>
</tr>
<tr>
<td>UV-3</td>
<td>0.2</td>
</tr>
<tr>
<td>UV-4</td>
<td>0.2</td>
</tr>
<tr>
<td>UV-5</td>
<td>0.2</td>
</tr>
</tbody>
</table>

Procedure. The compounds were screened for inhibition of generation of infectious virus was conducted on the UV compounds at concentrations up to 500 μM. The influenza virus, Influenza A H3N2, Brisbane/10/2007 strain was evaluated for virus inhibition. MDCK cells (Madin Darby canine kidney cell line) obtained from American Type Culture Collection (ATCC, Manassas, Virginia). Cells were cultured in UltraMDCK, supplemented with 2 mM L-glutamine, 1 μg/ml TPCK-treated trypsin and 100 U/ml penicillin, 100 μg/ml streptomycin in cell culture treated 24-well flat bottom plates at 37°C in a 5% CO2 incubator for 24 hr or until 80% confluent prior to assay. Cells were pretreated with compounds in a final concentration of 0.5% DMSO for 1 hr followed by addition of virus inoculums. Three wells per virus are saved for a virus-only control. Only medium is added in exchange for
compound in these wells, and virus is added after the initial 1 hr incubation. Three days later virus containing supernatants were collected and effect on reduction of virus yield are tested by assaying frozen and thawed eluates from each well for virus titer by serial dilution onto monolayers of MDCK susceptible cells. The 90% effective concentration (EC90), which is that test drug concentration that inhibits virus yield by 1 log10, is determined from these data.

Influenza In Vivo Study

UV-4 was administered as a free drug dissolved in acidic water. The compound was given at 100 mg/kg and 10 mg/kg by the oral route (intragastric via oral gavage - IG) twice daily. Balb/c mice received the compound for 10 days. Mice were infected with INFV A H1N1 (strain A/Texas) intranasally with ~5 LD50 30 minutes following the first iminosugar dose. Animals were monitored for 15 days. Animals were weighed once per day, and given health scores 2X per day. Animals displaying severe illness (as determined by 30% weight loss, extreme lethargy, ruffled coat, or paralysis) were euthanized.

Figure 5 shows effects of 10-day administration of UV-4 on survival of mice infected with influenza A H1N1.

Results: Animals receiving 100mg/kg and 10mg/kg BID showed a 90% survival rate, versus a 30% survival rate in control animals.

Conclusion: These results demonstrate that UV-4 can be used as a host-based antiviral drug to treat influenza A.

Iminosugar Safety Study

Methods and Discussion: BALB/c and C57/Bl/6 mice were given oral suspensions of UV-1, UV-4, UV-5, twice a day for seven days, in 100ul per mouse at 100 and 10 mg/kg (2mg and 0.2 mg/mouse, respectively) 8 hours apart for 7 days, and then monitored for weight loss and general health. After seven days of treatment, the mice did not show any significant signs of weight loss compared to the “vehicle only” control. The results of these experiments are in Figure 6.

When the BALB/c mice were treated with UV-5 at the highest concentration, they displayed signs of diarrhea, red urine, and a ruffled appearance although they did not show signs of weight loss. The C57/Bl/6 mice displayed these same symptoms but without the ruffled look.
These symptoms promptly ceased when treatment was done, and by day 11 (day 4 post compound treatment) the BALB/c mice in these groups looked very healthy. Conclusions: These compounds have shown to be relatively non-toxic in this mouse model and these concentrations of compound are deemed safe.

Second Influenza *In Vivo* Study

Figure 7 presents survival data after H1/N1 (Texas) virus challenge for mice treated with UV-4 versus control mice. UV-4 was administered to the treated mice as a free drug dissolved in acidic water by the oral route (intragastric via oral gavage - IG) 100mg/kg, TID for 10 days. The control mice received water orally, TID, instead of UV-4. Balb/c mice were used both for the treated mice and the control mice. Each mouse was microchipped for individual identity. Mice were infected with INFV A H1N1 (strain A/Texas) intranasally with ~1 LD90. Animals were monitored for 15 days. Animals were weighed once per day, and given health scores 2X per day. Animals displaying severe illness (as determined by 30% weight loss, extreme lethargy, ruffled coat, or paralysis) were euthanized. The endpoint was considered a death of the animal or a more than 30% weight loss. The studied mice include the following groups (10 mice per group):

1) 1 hr pre-treatment. These mice received their first UV-4 dose 1 hr before being infected with INFV A H1N1 (strain A/Texas).

2) 24 hr post-treatment. These mice received their first UV-4 dose 24 hr after being infected with INFV A H1N1 (strain A/Texas).

3) 48 hr post treatment. These mice received their first UV-4 48 hr after being infected with INFV A H1N1 (strain A/Texas).

4) 96 hr post treatment. These mice received their first UV-4 96 hr after being infected with INFV A H1N1 (strain A/Texas).

Results: Animals in the 1 hr pre-treatment and 24 hr post-treatment groups demonstrated 100% survival during the experiment’s duration, while mice in the 48 hr post-treatment and 96 hr post-treatment groups demonstrated 90% survival. The survival rate for the control mice was 30%.
Conclusion: These results demonstrate that UV-4 can be used as a host-based antiviral drug to treat and prevent influenza A.

* * *

Although the foregoing refers to particular preferred embodiments, it will be understood that the present invention is not so limited. It will occur to those of ordinary skill in the art that various modifications may be made to the disclosed embodiments and that such modifications are intended to be within the scope of the present invention.

All of the publications, patent applications and patents cited in this specification are incorporated herein by reference in their entirety.
WHAT IS CLAIMED IS:

1. A method of treating and/or preventing a disease or condition caused by or associated with a virus belonging to the Orthomyxoviridae family, the method comprising administering to a subject in need thereof an effective amount of a compound of the formula:

\[
\begin{align*}
W_1 & \quad \text{or a pharmaceutically acceptable salt thereof, wherein } R \text{ is } \\
W_2 & \quad \text{either selected from substituted or unsubstituted alkyl groups, substituted or unsubstituted cycloalkyl groups, substituted or unsubstituted aryl groups, or substituted or unsubstituted oxaalkyl groups; or wherein } R \text{ is } \\
W_3 & \quad \text{R}_1 \quad \text{is a substituted or unsubstituted alkyl group;} \\
W_4 & \quad \text{X}_{1-5} \text{ are independently selected from H, NO}_2, \text{ N}_3, \text{ or NH}_2; \\
W_5 & \quad \text{Y is absent or is a substituted or unsubstituted C}_1\text{-alkyl group, other than carbonyl; and} \\
W_6 & \quad \text{Z is selected from a bond or NH; provided that when } Z \text{ is a bond, } Y \text{ is absent, and provided that when } Z \text{ is NH, } Y \text{ is a substituted or unsubstituted C}_1\text{-alkyl group, other than carbonyl; and} \\
W_7 & \quad \text{wherein } W_{1-4} \text{ are independently selected from hydrogen, substituted or unsubstituted alkyl groups, substituted or unsubstituted haloalkyl groups, substituted or unsubstituted alkanoyl groups, substituted or unsubstituted aroyl groups, or substituted or unsubstituted haloalkanoyl groups.}
\end{align*}
\]
2. The method of claim 1, wherein each of \( W_1, W_2, W_3 \) and \( W_4 \) is hydrogen.

3. The method of claim 1, wherein \( R \) is selected from substituted or unsubstituted alkyl groups, substituted or unsubstituted cycloalkyl groups, substituted or unsubstituted aryl groups, or substituted or unsubstituted oxoalkyl groups.

4. The method of claim 1, wherein \( R \) is C6-C12 alkyl or oxoalkyl group.

5. The method of claim 1, wherein \( R \) is C8-C10 alkyl or oxoalkyl group.

6. The method of claim 1, wherein said administering comprises administering N-nonyl deoxyunjirimycin or a pharmaceutically acceptable salt thereof.

7. The method of claim 1, wherein said administering comprises administering N-(7-oxadecyl)deoxyunjirimycin or a pharmaceutically acceptable salt thereof.

8. The method of claim 1, wherein said administering comprises administering is N-(9-Methoxynonyl)deoxyunjirimycin or a pharmaceutically acceptable salt thereof.

9. The method of claim 1, wherein \( R \) is

10. The method of claim 9, wherein \( X_1 \) is NO\(_2\) and \( X_3 \) is N\(_3\).

11. The method of claim 9, wherein each of \( X_2, X_4 \) and \( X_5 \) is hydrogen.

12. The method of claim 1, wherein said administering comprises administering is N-(N-(4'-azido-2'-nitrophenyl)-6-aminohexyl)deoxyunjirimycin or a pharmaceutically acceptable salt thereof.

13. The method of claim 1, wherein the subject is a mammal.

14. The method of claim 1, wherein the subject is a human being.
15. The method of claim 1, wherein the virus is an Influenza virus belonging to the Influenza A, Influenza B or Influenza C genus.

16. The method of claim 15, wherein the virus is Influenza A virus.

17. The method of claim 16, wherein the virus is a H3N2 subtype of the Influenza A virus.

18. The method of claim 16, wherein the virus is a H1N1 subtype of the Influenza A virus.

19. The method of claim 18, wherein the compound is N-(9-Methoxynonyl)deoxynojirimycin or a pharmaceutically acceptable salt thereof.

20. The method of claim 19, wherein said administering prevents said disease or condition in the subject.

21. A method of inhibiting infectivity of a cell infected with a virus belonging to the Orthomyxoviridae family, the method comprising contacting a cell infected with a virus belonging to the Orthomyxoviridae family with an effective amount of a compound of the formula:

\[
\text{[Diagram of molecular structure]}
\]

or a pharmaceutically acceptable salt thereof, wherein \(R\) is either selected from substituted or unsubstituted alkyl groups, substituted or unsubstituted cycloalkyl groups, substituted or unsubstituted aryl groups, or substituted or unsubstituted oxaalkyl groups; or wherein \(R\) is
$R_1$ is a substituted or unsubstituted alkyl group;
$X_{1-5}$ are independently selected from H, NO$_2$, N$_3$, or NH$_2$;
Y is absent or is a substituted or unsubstituted C$_1$-alkyl group, other than carbonyl; and
Z is selected from a bond or NH; provided that when Z is a bond, Y is absent, and provided that when Z is NH, Y is a substituted or unsubstituted C$_1$-alkyl group, other than carbonyl; and
wherein $W_{1-4}$ are independently selected from hydrogen, substituted or unsubstituted alkyl groups, substituted or unsubstituted haloalkyl groups, substituted or unsubstituted alkanoyl groups, substituted or unsubstituted aroyl groups, or substituted or unsubstituted haloalkanoyl groups.
**Methods**

UV-4: Compound administered orally for 7 days

UV-5: Compound administered orally for 7 days

**Results**

UV-4: No significant weight loss

UV-5: No significant weight loss

No adverse events for both UV-4 and UV-5

**UV compounds are safe in vivo**
FIGURE 7: UV-4 THERAPEUTIC MODE VS. INFLUENZA H1N1

Mouse Survival

UV-4: 1hr pre-treatment
- UV-4, 100mg/kg, orally, TID
- Control, water, orally, TID

UV-4: 24hr post-treatment

UV-4: 48hr post-treatment

UV-4: 96hr post-treatment
INTERNATIONAL SEARCH REPORT

A. CLASSIFICATION OF SUBJECT MATTER
IPC(8) - C07D 211/30 (2010.01)
USPC - 546/248
According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED
Minimum documentation searched (classification system followed by classification symbols)
USPC: 546/248

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched
USPC: 546/242, 11, 184 (text search) Find search terms below

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)
PubWEST (PGPB,USPT,EPAB,JPAB), Google Scholar
iminosugars,orthomyxoviridae, influenza A, H3N2, H1N1, subtype, 5-deoxyxojirimycin, N-nonyl deoxyxojirimycin, N-(7-
exadecyl)deoxyxojirimycin, antiviral, anti-viral

C. DOCUMENTS CONSIDERED TO BE RELEVANT

<table>
<thead>
<tr>
<th>Category*</th>
<th>Citation of document, with indication, where appropriate, of the relevant passages</th>
<th>Relevant to claim No.</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>8, 10, 12, 19-20</td>
</tr>
</tbody>
</table>

* Special categories of cited documents:
  "A" document defining the general state of the art which is not considered to be of particular relevance
  "E" earlier application or patent but published on or after the international filing date
  "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
  "O" document referring to an oral disclosure, use, exhibition or other means
  "P" document published prior to the international filing date but later than the priority date claimed
  "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
  "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
  "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
  "&" document member of the same patent family

Date of the actual completion of the international search
06 October 2010 (06.10.2010)

Date of mailing of the international search report
14 OCT 2010

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